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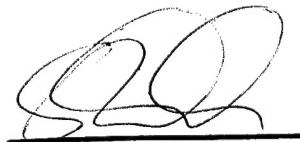
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Table of Contents

Front Cover	1
Report Documentation Page	2
Foreword	3
Table of Contents	4
Introduction	5
Body	5
Conclusions	8
References	10

Introduction

Chronic and acute stress has been associated with alterations in immune measures including Natural Killer (NK) cell activity (e.g., Herbert & Cohen, 1993). Healthy individuals with family histories of cancers have been shown to have lowered NK cell cytotoxicity (e.g., Strayer et al., 1984, 1986; Hersey et al., 1979). As NK cells are thought to serve an important function in immune surveillance against neoplastic cells (Trinchieri, 1990) it is possible that deficits in NK cell activity in individuals at familial risk for cancer may contribute to their heightened risk of developing the disease. It therefore becomes important to determine the causes of this lowered NK cell activity. Aside from heritable deficits in NK cell activity it is also possible that the higher levels of distress that have been found in women with family histories of cancer (e.g., Kash et al., 1992) may be partly responsible for their immune deficits. The present study explores the possibility that women with family histories of breast cancer may have higher psychophysiological reactivity and/or greater immunological sensitivity to stress than women without family histories of cancer. This is done using an experimental stressor paradigm that has been widely recognized in psychophysiological and psychoneuroimmunological research.

Body

Method

Subjects are exposed to two consecutive mental tasks that have been shown to affect psychophysiological reactivity (i.e., self-reported distress, cardiovascular changes, hormonal changes) as well as immune measures (i.e., NK cell activity) (e.g., Manuck et al., 1991; Stone et al., 1993; Zakowski et al., 1992). Self-reported distress, cardiovascular, hormonal and immunological measures are assessed before during and after stressor exposure at 15 to 30 minute intervals. Changes are assessed in response to the stressors and compared to resting baseline levels in order to determine the magnitude and duration of subjects stress reactivity and immunological sensitivity. These effects are compared to measures taken in response to a non-stressful control task. In addition, subjects complete questionnaires assessing family history of cancer, chronic distress, cancer-related symptoms of distress, etc. (see Measures). We hypothesize that: 1) Women with family histories of breast cancer show greater psychophysiological reactivity and immunological sensitivity than women without family histories of cancer. 2) Chronic stress, fear, and uncertainty associated with risk for cancer will partly account for subjects heightened psychophysiological reactivity and lower NK cell activity at baseline and in response to stress.

Measures

1. Psychobehavioral Study Measures. The standardized measures described briefly below were selected because of their possible relation to immune measures and because of their well established reliability and validity.

a. **Measures of chronic stress.** These questionnaires assess background levels of stress associated with daily life and specifically perceived risk of breast cancer. Their inclusion will permit assessment of group differences in distress and will enable us to examine the possible contribution of chronic stress to the psychophysiological and immunological responses to the laboratory tasks.

Life Experiences Survey (LES). The LES (Sarason et al., 1978) assesses the total number of life events and their impact, which has been reported to predict anxiety, depression, and psychological discomfort.

Brief Symptom Inventory (BSI). The BSI (Derogatis et al., 1982) with nine symptom dimensions and three global indices of distress has been used in previous studies of individuals with a family history of cancer.

The Perceived Stress Scale (PSS). The PSS (Cohen et al., 1983) assesses how unpredictable, uncontrollable, and overloading respondents find their lives, which may be related to immune function.

Impact of Event Scale (IES). The IES (Horowitz et al., 1979), assesses distress anchored to a specific stressor; in this case the threat of cancer.

Perceived Risk for Cancer. This face-valid questionnaire determines whether the women perceive themselves to be at increased risk for breast cancer.

b. **Measures of stress mediators.**

Courtault Emotional Control Scale. The CECS assesses emotional control and expressivity, which has been suggested by some investigators to differ in individuals with cancer.

c. **Other background measures.** These questionnaires assess variables, such as demographic variables and health habits that may affect physiological and immune measures.

Demographic questionnaire. The purpose is to obtain basic demographic information such as age, race, socio-economic status, etc.

Daily Habits Questionnaire (DHQ). The DHQ is a face valid instrument developed by us to assess sleep, physical activity, eating patterns, cigarette smoking, alcohol consumption, use of licit and illicit drugs, and menstrual cycles, variables that may affect immune function.

2. Measures of acute stress in response to the laboratory manipulations Reactivity to the laboratory stressors will be assessed at three levels. Psychological distress will be assessed by self-report questionnaires, cardiovascular reactivity will be assessed by continuous monitoring of heart rate (HR) and blood pressure (BP), and biochemical

measures of stress will include changes in levels of stress hormones.

a. Self-report measures

Visual Analog Scales (VASs). VASs (Cella et al., 1986) will be used to provide measures of subjects': distress associated with the assessment visit, current levels of emotional distress, venipuncture distress, which may be related to immune function.

The Profile of Mood States (POMS). The POMS (McNair et al., 1971) assesses current levels of emotional distress.

b. Cardiovascular measures. Blood pressure and heart rate is monitored at set intervals using an automated monitoring device.

c. Endocrine measures

Catecholamines in plasma samples collected at each assessment will be assayed by the CRC Core Laboratory at CUMC under the direction of Dr. Imperato-McGinley using classic HPLC techniques.

Cortisol in plasma samples will be assayed by the CRC Core Laboratory at CUMC using commercial radioimmunoassay kits with high reliability.

3. Immune measures. The immune measure of primary interest is NK cell activity, because of the published data indicating deficits in individuals with a family history of cancer (e.g., Strayer et al., 1984). In addition, psychoimmune studies have repeatedly documented that NK cell activity is sensitive to emotional distress (Herbert & Cohen, 1993).

Quantification of leukocyte subpopulations. Complete blood counts (CBC) with differential is done on the day of assessment by a laboratory using laser light scatter and enzyme cytochemistry. Flow cytometric quantifications of well established lymphocyte subsets (e.g., CD3+, CD4+, CD8+, CD19+, CD4+CD45RA+, CD4+CD45-), with particular emphasis on enumerating natural cytotoxic effectors based on phenotype (e.g., CD2, CD3, CD16, CD56), is accomplished with two color immunofluorescence techniques.

Effector Cells and Culture Conditions. Mononuclear cells are isolated from heparinized blood samples by standard Ficoll-Hypaque (Pharmacia, Piscataway, NJ) gradient centrifugation routinely yielding cells with greater than 95% viability, which is used fresh for the assays described here.

Natural killer cell activity. Natural killer cell activity is assessed in classic chromium release assays using the natural killer (NK) cell sensitive K562 erythroleukemia line (Bonavida et al., 1983).

Blastogenic responses and production of cytokines. Following classic methodologies, isolated mononuclear cells are stimulated with: the classic T cell dependent mitogens.

Results

As of December 31, 1995 (Month 12 of the study) we have recruited a total of 64 women with and without family histories of cancer who have participated in the first session of the study. Of those participants, 38 have completed the second experimental session. We therefore expect that, in line with our proposed Statement of Work, we will have completed the full study with a total of 100 subjects by the end of 1996. In accordance with our Statement of Work (see Appendix K of the grant) we will begin data processing and analyses by Month 17 of the study period (i.e., May 1996) and therefore cannot present results from the present study. It is routine procedure for a short-term experimental study such as this one to analyze the data once its collection has been completed so as not to unintentionally influence the rigorous experimental procedures by experimenters expectations based on preliminary findings. Based on initial observations it can be concluded, however, that our experimental paradigm is successful in eliciting the expected stress and immune effects that have previously been shown in the literature independent of family history of cancer. That is, the mental tasks elicit reliable increases in self-reported distress, as measured by visual analog scales, increases in heart rate and blood pressure, as well as a biphasic response curve in NK cell activity with an initial increase followed by a subsequent decrease in activity as expected based on previous literature (e.g., Schedlowski et al., 1993). This confirms our expectations and previously published data on the effects of experimental tasks on stress and immunological measures and confirms the methodological soundness of our research design. The quantitative analysis of response differences between women with family histories of cancer and women without family histories of cancer addressing our main hypotheses (see above) will be conducted in accordance with our Statement of Work.

Conclusions

Preliminary observations confirm previous data showing that when subjects are exposed to stressful mental tasks in a controlled laboratory setting increases in psychological and cardiovascular indices of distress as well as changes in immune function are seen. Analyses addressing the major study hypotheses will be conducted upon completion of the data collection. The final results from this study will determine the role of stress in the reduced NK cell acitivity in women at familial risk for cancer and will provide potential mechanisms by which stress may be partly responsible for these immune deficits. To date no other studies have addressed these issues in populations at risk for cancer and the findings from this study will provide important information on how women at familial risk for cancer respond to stress both psychophysiological and immunologically. The results will have important implications for designing stress-reducing interventions for women at familial risk for cancer to help them reduce the psychological impact of stressful events and as a consequence to attempt to attenuate the potentially deleterious effects of stress on the immune system in these women. The findings will also lead to further studies refining previous methodology in order to determine in greater detail the sources of distress in this population so as to be able to target more specific stressors for intervention purposes.

Finally, this study will contribute to a greater awareness of the importance of psychological issues in cancer risk for both researchers and clinicians.

References

- Bonavida, B., Bradley, T.P., Grimm, E.A. Frequency determination of killer cells by a single-cell cytotoxic assay. 1983 Methods in Enzymology, 93:270-280.
- Cella DF, Perry SW. Reliability and concurrent validity of three visual-analogue mood scales. Psychol Rep 1986; 59:827-833.
- Cohen S, Karmarck T, Mermelstein R. A global measures of perceived stress. Journal of Health and Social Behavior 1983; 24:385-296
- Derogatis LR, Spencer P. The Brief Symptom Inventory (BSI) Administration Scoring and Procedures Manual-I. Baltimore: copyrighted manuscript, 1982.
- Herbert TB, Cohen S. Stress and immunity in humans: A meta-analytic review. Psychosomatic Medicine 1993; 55:364-379.
- Hersey P, Edwards A, Honeyman M, McCarthy WH. Low natural killer-cell activity in familial melanoma patients and their relatives. Brit J Cancer 1979; 40:113-122.
- Horowitz M, Wilner N, Alvarez W. Impact of Event Scale: A measure of subjective stress. Psychosom Med 1979; 41:209-218.
- Kash KM, Holland JC, Halper MS, Miller DG. Psychological distress and surveillance behaviors of women with a family history of breast cancer. J Natl Cancer Inst 1992; 84:24-30.
- Manuck SB, Cohen S, Rabin BS, Muldoon MF, Bachen EA. Individual differences in cellular immune response to stress. Psychological Science 1991; 2:111-115.
- McNair DM, Lorr M, Droppleman LF. Manual: Profile of Mood States. San Diego:Education and Industrial Testing Service, 1971.
- Sarason IG, Johnson JH, Siegel JH. Assessing the impact of life changes: Development of the life experiences survey. Journal of Consulting and Clinical Psychology 1978; 46:932-946.
- Schedlowski M, Jacobs R, Stratman G, Richter S, Haedicke A, Tewes U, Wagner T, Schmidt R: Changes of natural killer cells during acute psychological stress. Journal of Clinical Immunology 1993; 13:119-126.
- Stone AA, Valdimarsdottir HB, Katkin ES, Burns J, Cox DS, Lee S, Fine J, Ingle D, Bovbjerg DH. Effects of mental stressors on mitogen-induced lymphocyte responses in the laboratory. Psychology and Health 1993; 8:269-284.
- Strayer DR, Carter WA, Mayberry SD, Pequignot E, Brodsky I. Low natural cytotoxicity of peripheral blood mononuclear cells in individuals with high familial incidences of cancer. Cancer Research 1984; 44:370-374.
- Strayer DR, Carter WA, Brodsky I. Familiar occurrence of breast cancer is associated with reduced natural killer cytotoxicity. Breast Cancer Research Treatment 1986; 7:187-192.
- Trinchieri G. Biology of natural killer cells. Advances in Immunology 1990; 47:187-376.

Zakowski SG, McAllister CG, Deal M, Baum A: Stress, reactivity and immune function. Health Psychology
11:223-232, 1992